Effective of Red Ginseng Extract on Ameliorate Adipogenesis 3T3-L1 by Decreasing P38 and Erk Phosphorylation

Seon Mi Ko, Seung-Mi Hwang, Dongjoong Im, Kyung-Min Choi*, Gi-Beum Kim*

Institute of Jinan Red Ginseng, 41 Hongsamhanbang-ro, Jinan-eup, Jinan-gun, Jeonbuk, 55442 Republic of Korea

*Corresponding author: Gi-Beum Kim, Institute of Jinan Red Ginseng, 41 Hongsamhanbang-ro, Jinan-eup, Jinan-gun, Jeonbuk, 55442 Republic of Korea, E-mail: kgb70@jbnu.ac.kr
Kyung-Min Choi, Institute of Jinan Red Ginseng, 41 Hongsamhanbang-ro, Jinan-eup, Jinan-gun, Jeonbuk, 55442 Republic of Korea, E-mail: kyungmc@ijrg.re.kr

Abstract
This study examined the anti-obesity effect and mechanism of action of red ginseng extract (RG) in 3T3-L1 cells. To measure the levels of lipid accumulation, we observed expression of genes and proteins, which is associated with adipocyte differentiation in 3T3-L1 cells. Eight days following induction for differentiation with MDI-medium and simultaneously with the tested RG, our study found significantly reduced (p < 0.05) intracellular triacylglycerol accumulations of 3T3-L1 cells by doses dependent manner. RG decrease adipogenic markers, C/EBPα, C/EBPβ and PPARγ, which is a key regulator of adipocyte differentiation. In this study showed that C/EBPβ is regulated RG decreased the phosphorylations of p38 mitogen-activated protein kinase (MAPK) and extracellular signal–regulated kinases (ERK). These results suggest that the anti-adipogenic effect of RG in 3T3-L1 pro-adipocyte could be associated with the inhibition of adipogenic transcription factors by regulations with p38 MAPK and ERK phosphorylation.

Keywords: Red ginseng extract: Anti-obesity effect; 3T3-L1 cells; p38; Mitogen-activated protein kinase; Extracellular signal–regulated kinases

Introduction
Botanical medicines have been applied for the treatment of various human diseases with thousands of years of history in Asia and are sharing a large market in the form of drugs, dietary supplements, and foods. In the west, botanical medicines are categorized as complementary/alternative medicines, dietary supplements, or foods. Ginseng, referred to as the root of Panax ginseng Meyer (Araliaceae), is one of the most valuable medicinal plants, particularly in Korea, China, and Japan[1]. Ginseng has been used as a valuable tonic and for the treatment of various diseases[2]. The pharmacological properties of ginseng are mainly attributed to ginseng saponins, commonly called ginsenosides, the major and bioactive constituents[3]. With the development of modern chromatography, there are more 40 ginsenosides such as ginsenosides Rb1, Rb2, Rg1, Rd, and Re identified from ginseng up to date Botanical characteristic[3,4].

Ginseng is a well-known medicinal plant used in traditional Oriental medicine. It has long been known to possess a variety of beneficial pharmacological effects, such as enhancement of immunity, inhibition of inflammatory response, and hyperlipidemia and diabetes prevention[5-8]. Red ginseng has been heat steamed and dried. As a consequence of this process, red ginseng undergoes certain biochemical changes[9]. However, some studies to show that it possesses pharmacological properties like immunomodulatory and anti-cancer effects[10,11].

Obesity is one of the greatest public health problems and major risk factors for serious metabolic diseases and significantly increases the risk of premature death[12,13]. Various anti-obesity medications have been developed, including orlistat, lorcaserin, and sibutramine. But, these medications can limit long term use, those have serious side effects such as steatorrhea, high blood pressure, constipation, insomnia, and hepatotoxicity[14,15]. Thus, novel preventive and therapeutic options with greater efficacy and fewer side effects are needed.

The MAPK/ERK pathway regulates the control of cell survival and proliferation. The MAPK/ERK signaling cascade is organized hierarchically into a three-tiered module composed...
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Materials and Methods

Cell culture
3T3-L1 preadipocytes were purchased from American Type Culture Collection (ATCC). 3T3-L1 preadipocytes were propagated in high glucose DMEM supplemented with 10 % (v/v) FCS (full name) and 1 % penicillin-streptomycin at 37 °C in a humidified atmosphere having 5 % CO2. Differentiation was induced in confluent cells by replacing DMEM (Dulbecco Modified Eagle Medium) with differentiation media. Differentiated medium was contained 10 mg/l insulin, 0.5 mM IBMX (3-isobutyl-1-methylxanthine) and 0.25 μM dexamethasone for 48 hrs.

Real time-PCR analysis
Total RNA was extracted by using easy-spin™ (DNA free) Total RNA Extraction Kit (intron (??)). One microgram of total RNA was subjected to first strand c DNA synthesis with oligo (deoxythymidine) primers and PrimeScriptTM RTase (TaKaRa).

The reaction mixture (contain Oligo dT primer, dNTP mixture, and template RNA) was incubated 5 min at 65 °C and then on ice. Prepare reaction mixture add Prime Script Buffer, RNase inhibitor and Prime Script RTase for 1 cycle at 30 °C for 10 min, 42 °C for 60 min, 95 and 4 °C for 10 min. The PCR reactions were performed as follows: 5 min at 94 °C for 5 min and 4 °C. PCR reaction was used to AccuPower® PCR Premix (Bioneer) 30 cycles of 15 sec at 94 °C, 40 sec at 55 °C, and 45 sec at 72 °C; and a final extension of 7 min at 72 °C. The PCR amplicons were then electrophoresed in 1 %-agarose gels containing 40 mM Tris, 20 mM glacial acetic acid and 2 mM EDTA buffer.

Western blotting
Cells were washed twice with cold PBS and were harvested in 200 μl RIPA buffer (150 mM NaCl, 1.0 % IGE-PAL® CA-630, 0.5 % sodium deoxycholate, 0.1 % SDS, 50 mM Tris, pH 8.0) and protease inhibitor (1 mM PMSF, 1 mg/l leupeptin, 1 mg/l aprotinin, and 10 mg/l phosphatase inhibitor cocktail). Cell lysates were centrifuged at 13,000 g for 30 min at 4 °C. The protein content of the supernatant was measured by using the BSA protein assay kit (Pierce Chemical Company). Equal amounts of protein for each sample were applied to 10 % SDS–PAGE gels and were electrophoretically transferred to polyvinylidene fluoride membranes. After electrophoresis the nonspecific binding sites in the membranes were blocked with 5 % nonfat dry milk in Tris-buffered solution (15 mM Tris-150 mM NaCl, pH 7.4) at room temperature for 1 h. After blocking, the membrane was incubated with anti-adipsin, anti-C/EBPα, anti-C/EBPβ, anti-PPARγ, anti-phospho-ERK1/2, anti-ERK1/2, anti-phospho-p38, anti-p38 and anti-β-actin antibodies at 4 °C overnight. Thereafter, the membrane was incubated with the secondary peroxidase-conjugated anti-rabbit or anti-mouse IgG at 37 °C for 90 min. The bands were detected by using an enhanced chemiluminescence kit (Pierce Chemical Company, Rockford, IL)

Statistical analysis
Data are expressed as means ± standard errors of the mean (SEM). Differences between groups were evaluated by analysis of variance (ANOVA) with the Bonferroni post hoc test or by calculation of Spearman’s rank correlation coefficient, as appropriate, using Prism 5.03 (Graph Pad Software Inc., San Diego, CA, USA). Statistical significance was set at p < 0.05.

Results
RG inhibits lipid accumulation in 3T3-L1 pre-adipocytes
We first determined the effects of RG on cell viability. Preadipocytes 3T3-L1 cells were treated with 0, 0.0039, 0.0078, 0.0156, 0.0312, 0.0625, 0.125, 0.25, 0.5, 1, 2, 4 mg/ml for 24 hrs as the figure 1A, cell viability was 99.3, 97.2, 92.4, 90.5, 90.3, 83.4, 18.9, 7.3, 7.0 %. The cytotoxic effect of RG (up to 125 μg/ml) in pre-adipocytes was determined using MTT assay at different doses. There was no significant decrease in viability at the tested concentration (figure 1A). To determine the effects of RG on the adipocyte differentiation, 3T3-L1 pre-adipocyte were incubated until 2-day post-confluence (day 0) and treated with RG at a concentration of 15.6, 31.3, 62.5, 125 μg/ml for 8 days during a standard adipogenic induction. RG was determined to significantly inhibit lipid accumulation in a dose-dependent manner through the microscopic examination of Oil Red O-stained lipid droplets and through the spectrophotometric quantification of the extracted stain, indicating the inhibition of adipocyte differentiation in 3T3-L1 (figure 1B).

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Expression of C/EBPα, C/EBPβ and PPAR γ are inhibited by RG

Adipocyte differentiation result in a series of programmed alternations of specific genes. Adipogenic is related by adipokines such as PPARγ, C/EBPα, and C/EBP β which are known to be critical activators of adipogenesis. In the fully differentiated adipocyte, RG treatment of 15.6, 31.3, 62.5 and 125 μg/ml decreased C/EBP α and by approximately 0.8, 48.1, 56, and 61.8 % compared to the control. C/EBP β is decreased by RG -2.4, 22.8, 23.6, 48.8, and 71 % compared to the MDI treated only. To determine the change of expression level of protein and mRNA, we accomplished western blotting and RT-PCR. The values above the figures represent relative density of the bands normalized to β-actin. C/EBPα is decreased by RG at 15.6 μg/ml, and C/EBP β protein level is decreased at 125 μg/ml at 4 days. At the same time, there proteins, which are C/EBP α and C/EBP β, levels are reduced by RG in a dose-dependent manner (Figure 2).

Figure 1: The cytotoxic effect of RG in preadipocytes by MTT assay test in various doses. A; cell viability, B; staining intensity.

Figure 2: The protein expression of CCAAT enhancer binding protein (C/EBP) β and alpha in 3T3-L1 cells incubated with RGM. The 3T3-L1 cells were treated with different concentrations of RGM (0, 15.6, 31.3, 62.5, 125 μg/ml) for 4 day. The upper panels are RT-PCR of C/EBP alpha and beta(A), flod change of C/EBP alpha and beta. The lower panels are western blots of C/EBP alpha, beta, and PPAR gamma.

Figure 3: The protein expression of adepsin and p21 in 3T3-L1 cells incubated with RGM. The 3T3-L1 cells were treated with different concentrations of RGM (0, 15.6, 31.3, 62.5, 125 μg/ml) for 4 day. The left panels are western blot of adepsin and p21 (A), and the right panels are fold change of C/EBP alpha and beta.
Expression of Adipsin and p21 inhibited by RG

To determine the effect of the RG on expression level of proadipogenic marker, adipsin and p21, western blot assay is performed. p21 and adipsin expression level is reduced by RG, dose dependent manner. Adipsin and p21 are decreased by RG at 62.5 μg/ml and 15.6 μg/ml respectively. (Figure 3), RG treatment of 15.6, 31.3, 62.5 and 125 μg/ml decreased Adipsin by approximately -7.6, -13.7, -11, 11.4, and 35.4 % compared to the control. p21 protein expression level is reduced by RG, 5.9, 12.4, 7.9, -0.7 and 9.0 % when treated by RG 15.6, 31.3, 62.5 and 125 μg/ml respectively.

The protein expression of p-p38, p38, p-ERK, ERK and β-actin in 3T3-L1 cells incubated with RG

Adipocyte differentiation and lipolysis were closely related with MAPK proteins. Specially, previous study is reported that p38 and ERK were regulated adipogenesis[18,19]. Activated p38 and ERK expression level were decreased by RG in a dose dependent manner. Activated p38 expression level was inhibited by RG treatment (15.6, 31.3, 62.5 and 125 μg/ml) 5.9, 1.9, 1.6, 11.7, 19.7, and 24.1%. Also, activated ERK is decreased 0, -16, -22.3, -5.6, 24.5 and 45.1 % by RG 15.6, 31.3, 62.5 and 125 mg/ml respectively in similar tendency with p38 reduction. (Figure 4)

Figure 4: The protein expression of p-ERK, ERK, p-p38, p38 and β-actin in 3T3-L1 cells incubated with RG. The 3T3-L1 cells were treated with different concentrations of RG (0, 3.8, 7.8, 15.6, 31.3, 62.5 μg/ml) for 4 days.

Discussion

Obesity is caused by overabundant fat accumulation and an excess number of adipocyte in adipose tissue and associated with metabolic disease, hyperglycemia, insulin resistance, and cardiovascular diseases. Lipid differentiation is related with adipogenic transcription factors, lipidogenesis factors and adipogenic hormone. One of the key regulator is MAPK signal[20]. MAPK signal protein, such as ERK and p38, are regulates cell differentiation and also cell proliferation. Schild, et al. is reported that p38 regulates PPAR γ[21]. Decrease of PPAR γ activity lead to reduction of adipsin and p21, adipogenic marker[22].

In the previous study, red ginseng extracts regulated energy expenditures in adipocyte tissue[23]. RG which is remained extract of red ginseng has anti-inflammatory effects, anti oxidative effects and hypoglycemic effect as red ginseng extract[11,24]. In this study, we confirmed that red ginseng extract inhibition of adipogenesis in 3T3-L1, preadipocytes cells.

To play a key role in controlling the differentiation of preadipocytes into adipocyte, a cascade of transcription factor expression and activation is well demonstrated. The importance of C/EBP α, β and δ and PPAR γ is supported by the finding that white adipose tissue formation is suppressed by gene deletion in C/EBP β/- δ/- or C/EBP α/- or PPAR γ/- knockout mice[15,26]. Adipocyte proliferation and differentiation is regulated by adipogenic transcription factors such as PPAR γ, C/EBP α, and C/EBP β and the regulator of lipogenic genes SREBP1 (for sterol-regulatory-element binding protein 1)[27-29]. RG is reduced the expression of adipogenic transcription factor, PPAR γ, C/EBP α and C/EBP β. This reduction of transcription factors is led by decline of p38 and ERK activation. Bak, et al. is reported RG concerned with p38 signal in macroage inflammatory reaction[11]. This report showed that RG is decreased p38 and ERK activation too in differentiation of pre-adipocyte 3T3-L1. C/EBP β and PPAR γ was regulated P38 ad ERK activation (Figure 5). Down regulation of C/EBP β and PPAR γ caused inhibition of adipogenesis in 3T3-L1 cells.

Figure 5: Inhibition pathway of RG in 3T3-L1 adipocyte differentiation

Conclusions

Our data demonstrate that RG inhibition of adipocyte differentiation through to regulation of MAPK activation. To regulation of MAPK regulate, draw reduction of adipogenic transcriptional factor (C/EBPα, C/EBPβ) and PPAR γ) and adipogenic makers (adipsin, p21) expression. Via these mechanisms RG is inhibition to accumulation of lipid in adipocyte. Our data suggest that RG can be used in a treatment of adipocyte.
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