Neuroprotective Effect of Rosuvastatin on CA1 & CA3 Regions of Hippocampus in High Fat Diet and Stress Induced Rats

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Abstract

Back ground: Prolonged stress and diet rich in cholesterol is known to cause memory impairment and cause disruption of neuronal network in hippocampus. In the present study, we evaluated the neuroprotective effect of rosuvastatin HMG CoA reductase inhibitor on stress induced and high fat diet fed rats given in combination.

Materials & methods: Forty eight adult male wistar rats were randomly assigned into eight groups. (N=6) control group received normal diet (chow diet), second group received only high fat diet (HFD), third group received only stress (STS), fourth group high fat diet and stress (HFD+STS), fifth group control + rosuvastatin 10mg/body wt, sixth group control + rosuvastatin 20mg/body wt, seventh group received high fat diet + stress and treated with rosuvastatin 10mg/body wt (HFD+STS+ROS 10mg), eighth group was treated with rosuvastatin 20mg/body wt (HFD+STS+ROS 20mg). Spatial memory was assessed by morris water maze. Rats were sacrificed at 96th day; brains were removed and processed for histological studies using cresyl violet staining. Neuronal population of CA1 & CA3 region was quantified.

Results: A significant (p < 0.01) increase in the neuronal population in the sub regions of hippocampus and improvement of spatial memory (CA1: 32±1.95, CA3: 22±1.5, latency to enter the target quadrant LT: 6.05±0.9sec, time spent in target quadrant TST: 2.46±1sec) was seen in rats treated with rosuvastatin 20mg/kg.body.wt (HFD+STS+ROS 20mg) compared to high fat diet and stress (HFD+STS): (CA1: 28±1.33, CA3: 18±1.47 & LT: 7.67±0.77 sec, TST: 10±1.37 sec. The rats treated with 10mg/kg.body wt (CA1: 30±1.83, CA3: 22±1.37) did not show any significance compared to HFD+STS group.

Conclusion: The results clearly demonstrate that rosuvastatin 20mg/kg.body.wt was able to prevent hippocampal neuronal loss in CA1 and CA3 regions and also enhanced learning and memory abilities. These findings support the view that rosuvastatin have neuroprotective action.

Keywords: Hippocampus; Rosuvastatin; Morris water maze

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Introduction

Cholesterol is the main constituent of the high fat diet. Pre-clinical studies have shown that dietary cholesterol can be the cause for neurodegenerative diseases and dementia (Alzheimer’s disease)[1,2]. Animal studies showed that high fat diet impairs hippocampal neurogenesis in male wistar rats[3] and epidemiological studies support the view that dietary cholesterol is one of the causes for Alzheimer’s disease and vascular dementia[4].

Prolonged stress alters the possible mechanisms like hypothalamic-pituitary-adrenal axis, neurotransmitters, decreases brain cell proliferations and brain corticosterone level during prenatal development[5]. Animal studies and clinical studies have demonstrated the stress effects cognitive tasks and it is one of the risk factor for dementia. Reports have suggested that chronic stress have reduced BDNF(brain derived neurotropic factor) values[6]. Recent findings suggested that combination of high fat diet and stress have produced extensive atrophy of dendrites of pyramidal neurons in hippocampal area[7].

Rosuvastatin is a hypolipidemic drug (statin) which acts by inhibiting rate limiting HMG-CoA reductase. Recent reports suggest that rosuvastatin exert additional pleiotropic activities that are independent of its hypolipidemic activity which includes antioxidant activity, anti-inflammatory activity, improvement of endothelial function, and stabilization of atherosclerotic plaques[8]. These findings have led to the speculation that rosuvastatin may have potential therapeutic implications in various neurological disorders. Investigations addressing the relationship between dementia and statins are mixed. Findings from cross-sectional studies suggested a protective role for statins in dementia but few prospective designed and cohort
studied, showed conflicting results[9,10].

**Aims and Objectives**

The aim of the present study is to explore the neuroprotective action of rosvuastatin in CA1 and CA3 regions of hippocampus when high fat diet and prolonged stress given to rats in combination.

**Materials and methods**

**Animals**

Inbred male Wistar strain weighing 120-150g were selected and procured from central animal house, Mamata Medical College Khammam. Six rats were housed in propylene cages (22.5 × 35.5 × 15 cm) maintained in 12 hours light and dark cycle in temperature and humidity controlled environment, and were fed with standard food pellet and water ad libitum. The experimental protocol was approved by the institutional animal ethical committee. (IAEC/DP-05/C16). All the experimental procedures were carried out during 10 am to 4 pm. Animals were randomly assigned into three groups of seven each.

Group 1: (control) Fed with standard chow diet for three months.
Group 2: Fed with high fat diet for three months (HFD)
Group 3: Fed with high fat diet and received stress for 21 days (HFD + STS)
Group 4: Fed with high fat diet and treated with 10 mg/kg.body wt (HFD + ROS 10 mg)
Group 5: Fed with high fat diet and treated with 20 mg/kg.body wt (HFD + ROS 20 mg)
Group 6: Fed with high fat diet and received stress and treated with rosvuastatin 10 mg/kg.body wt (HFD + STS + ROS 10 mg)
Group 7: Fed with high fat diet and treated with 20 mg/kg.body wt (HFD + STS + ROS 20 mg)

**Method of inducing hyperlipidemia:** Hyperlipidemia has been induced by feeding with cholesterol-rich diet for 3 months. Deoxycholic acid (5g) was mixed thoroughly with 700g of powdered rat chow diet. Simultaneously cholesterol (5g) was dissolved in 300g warm coconut oil. This oily solution was added slowly into the powdered mixture to obtain a soft homogenious cake. This cholesterol-rich diet was made into pellets of about 3g each and given to the animals[11].

**Induction of stress to the rat’s protocol:** Male wistar rats were placed in a wire mesh restrainer for 21 days, daily 6 hours, from 69th to 90th day. The restrainer was made up of a wooden base to which stainless steel wire mesh was hinged. A pad lock and a small pin was used to secure the rat. The dimensions were 8 cm (Length) x 4cm (Breadth) x 4 cm (Height). This type of wire mesh restrainer can only restrict the animal movement without any uneasiness, pain or suffocation[12].

**Treatment with rosvuastatin:** Rosvuastatin was obtained from Pfuger Company Mumbai. Groups 5, 6, 7, 8 rats were treated with 10mg/kg body weight and 20mg/kg body weight rosvuastatin for 14 days from 76th - 90th day[13].

**Morris water maze test:** Spatial memory of the rats was tested on 90th day by using Morris water maze. The water maze consists of a circular tank of 1.80 m diameter and 75 cm depth. The pool was filled with water and maintained at a temperature of 24-26°C to a depth of about 50 cm. It was divided into four quadrants and an escape platform of size 4"×4" was hidden approximately 2cm below the water surface in the target quadrant. Water in the pool was made opaque by adding milk just before the experiment. Permanently positioned distinctive objects were placed for facilitating spatial orientation of the animal. Positions of the cues were kept unchanged throughout the period of experiment. The rats were trained in the water maze in 10 sessions on 5 consecutive days, two sessions on each day. Each session consists of four trials. In each trail time taken to reach hidden platform was recorded. If the rat was unable to find the platform within two minutes, the training session was terminated and a maximum score of two minutes was assigned.

Weight four hours after the last session, rats were subjected to a probe trail. This session was for 30 in which the hidden platform was removed. Here time taken to reach the target quadrant and time spent by the rats in search of the platform was measured. Greater latency to reach the target quadrant and less time to spend in the target quadrant suggests memory impairment[14].

**Weight gain** Weight of the animals was measured at the beginning of the study and at the end (after 3 months) to calculate the net weight gained.

**Lipid profile** Blood samples were obtained by retro-orbital puncture, serum cholesterol and triglycerides were estimated on 96th day[15].

**Cresyl violet staining procedure** The animals were profoundly anesthetized with ether and fixed to dissection board. The chest cavity was opened and heart was exposed. About 15ml of 0.9% heparinized saline was perfused through left ventricle at the rate of 1ml/min followed by 250ml of 10% formalin. The animal was decapitated and the brain was isolated. 5-6mm thick coronal sections of brain tissue were made and kept in 10% formalin for 48 hours (post fixation). Paraffin blocks were made and sections of 5 microns thickness were cut from the mid dorsal hippocampal level, using a rotary microtome. Sections were selected and mounted serially on air dried gelatinized slides. Tissues were processed through different grades of alcohol (50%, 70% for 24 h, 90%, and 100% for 12 h) and were immersed in xylene for 1-2 h.

**Preparation of Cresyl violet stain (0.1%)** 100mg of Cresyl violet was dissolved in 100ml of distilled water. To this 0.5 ml of 10% Acetic acid was added to give a pH of 3.5 to 3.8. The stain was filtered before use.

**Quantification of cells** In each hippocampal section, cornua amonis(CA1 and CA3; 250 µm length) was selected using an oculomicrometer. The number of viable neurons were counted and averaged under (40X) magnification (Magnus, Olympus Pvt. Ltd. New Delhi, India). Darkly stained, shrunken and cells with fragmented nuclei were excluded from the count. To avoid manual bias ten sections from each rat were considered. The cell counts were expressed as the number of cells per unit length of the cell field (cells/250 µm)[16].
Statistical analysis

The data were expressed as mean ± SE. The significant differences among the groups were assessed using one way analysis of Variance (ANOVA) followed by Bonferroni multiple comparison test in Graph Pad in Stat (GPIIS) software, version 1.13. p values < 0.05 were considered as significant

Results and Statistical Analysis

Weight of the animals was measured during the study period (before and after three months). There was significant increase in the body weight of the animals fed with only high fat diet (HFD: 264± 6.90, (p<0.001) and high fat diet + stress (HFD+STS: 172 ± 9.61(p<0.01), when compared to control group animals (C: 149±7.36).

Significant decrease in body weight was observed in rats treated with rosuvastatin 10mg (HFD+STS+ROS 10mg/ kg.body.wt:152 ± 3.76, HFD+ROS10mg/kg.body.wt:243±7.08 p<0.01) compared to HFD+STS group and HFD groups. In rats treated with 20mg rosuvastatin also there was very significant decrease in body weights (HFD+STS+ROS 20mg/kg.body.wt:145 ± 4.47, HFD+ROS20mg/kg.body.wt:235±13.8 p<0.001) compared to (HFD+STS and HFD groups).

After three months serum cholesterol (cho) and triglyceride (tri) levels were elevated in both groups namely high fat diet (HFD) group (cho: 166±4.64, tri: 149 ±6.31), (p<0.001) and HFD + STS (cho: 92.2±3.43, tri: 101 ± 5.85), (p<0.01) as compared to the control group (cho: 80.7±5.79, tri: 88.2± 5.96).

In groups treated with rosuvastatin 20 mg/kg.body.wt and 10mg/kg.body.wt there was significant decrease in cholesterol and triglycerides. In rosuvastatin 10mg treated groups the significance was p<0.01 (HFD+ROS10mg/kg.body.wt: cho 154±4.92, tri 137.8±5.12, HFD+STS+ROS10mg/kg.body.wt cho 82.3±2.7, tri: 88.3±4.08) compared HFD and HFD+STS group. In Rosuvastatin 20mg treated groups the significance was p<0.001 (HFD+ROS20mg/kg.body.wt:cho139±3.76, tri 124.8±4.5, HFD+STS+ROS20mg/kg.body.wt cho 80.5±6.92, tri: 85.5±3.39) compared to HFD and HFD+STS groups.

Behavioural test (Morris water maze): Latency to enter the target quadrant

This was done on 96th day, twenty four hours after the 10th training session. Compared to control group (C: 3.32 ± 0.76 sec, high fat diet group HFD: 4.66 ± 0.84 sec took longer time (p<0.05) and high fat diet plus stress group HFD+STS: 7.67± 0.77sec took significantly (p<0.001) more time to reach the target quadrant (probe) suggesting memory impairment.

Rats treated with 10mg rosuvastatin took (HFD+ROS-10mg/kg.body.wt) 3.95 ± 0.75 sec and (HFD+STS+ROS 10mg/kg.body.wt) 7.07±0.56 sec which did not show any significance (p>0.01) significantly less time to enter the target quadrant (HFD+ROS20mg/kg.body.wt) 3.31±0.4 sec and (HFD+STS+ROS 20mg/kg.body.wt) 6.05±0.91 sec compared to HFD+STS group. The above results imply that only rosuvastatin given in higher dose 20mg have improved the spatial memory. (Figure 1)

Figure 1: Bar graph showing the latency to enter target quadrant (sec). Bars represent mean ± SEM (control vs HFD + STS, p < 0.001, control vs HFD** p < 0.05) (HFD + ROS 20 mg vs HFD*** p < 0.01) (HFD + STS + ROS 20 mg vs HFD + STS$$ p < 0.01).

Time spent in the target quadrant

High fat diet plus stress induced rats (HFD+STS: 10 ± 1.37 sec) spent significantly (p<0.001) lesser amount of time and rats fed with only high fat fed group took less time (p<0.05, HFD: 20.2 ±2.46 sec) in the target quadrant (probe) in search of the missing platform compared to control group (C: 24±3.01sec). This shows that both stress and high fat diet in combination produces more memory deficits.

The spatial memory of rats treated with rosuvastatin 20mg was significantly improved in (HFD+STS+ROS 20mg/ kg.body.wt: p<0.01, 14.3± 1.62 sec) and took longer time in search of the missing platform but (HFD+ROS: 10mg 13.2±1.51sec) group did not show any difference compared to the high fat diet plus stress group HFD+STS.

The spatial memory of the rats treated with 20 mg rosuvastatin in HFD +ROS 20 mg/kg.body.wt 24.5±1.77 sec took significantly less time p<0.01 but rats treated with 10mg (HFD+ROS 10 mg/kg.body.wt 19.1±2.17 sec) did not show any significant compared to HFD group. This shows that rosuvastatin given in higher dose enhances spatial memory. (Figure 2)
Histology hippocampus (cresyl violet staining)
The number of viable neurons of CA1 and CA3 regions of hippocampus showed significant decrease in High fat diet +stress group (HFD+STS: CA1:28±1.33, CA3:18± 1.47,p < 0.001) and only high fat diet fed group (HFD: 30.2±2.04, 24±1.97 p < 0.05) compared to control group (C: 34± 1.47, 27± 0.98).

The number of viable neurons in CA1 and CA3 regions in the rats treated with rosuvastatin 20mg group (HFD+STS+ROS 20mg/kg.body.wt:CA1:31± 1.75, CA3: 22± 2.12; HFD+ROS20mg/kg.body.wt: CA1: 30± 2.04, CA3:26.7±1.21 (p < 0.01) has significantly increased when compared to (HFD+STS) and HFD groups. The group which was treated with rosuvastatin 10mg (HFD+STS+ROS10mg/kg.body.wt CA1:29±0.33, CA3: 20± 0.44) (HFD+ROS10mg/kg.body.wt: CA1:30±1.8, CA3:22.3±1.37) did not show any significance when compared with HFD+STS group and HFD group. This implies the neuroprotective action of rosuvastatin in higher dosage 20mg/kg.body.wt (Figure 3, 4, 5 & 6).

Figure 3: Bar graph showing the no of surviving neurons in CA3 region of hippocampus. Bars represent mean ± SEM (control vs HFD+STS ### p < 0.001, control vs HFD** p < 0.01) (HFD+ROS 20 mg vs HFD* p < 0.01) (HFD + STS +ROS 20 mg vs HFD + STSp < 0.01).

Figure 4: Bar graph showing the no of surviving neurons in CA1 region of hippocampus. Bars represent mean ± SEM (control vs HFD + STS** p < 0.001, control vs HFD** p < 0.01) (HFD + ROS 20 mg vs HFD* p < 0.01) (HFD + STS +ROS 20 mg vs HFD + STSp < 0.01).

Discussion

The present study indicates that rosuvastatin in higher dosage 20mg has neuroprotective action on the neurons of ca1 and ca3 regions of hippocampus when compared to rats fed with high dietary fat and induced to stress.

In behavioural models (Morris water maze) a marked decrease in escape latency, as compared to first exposure denotes normal learning ability. Increase in the time spent by the animal in search of the missing platform in the target quadrant and decrease in time taken to reach the target quadrant gives us the successful retention of learned memory. In the present study rats treated with rosuvastatin 20mg has taken less time to reach the target quadrant and spent significantly more time in the target quadrant in the search of the missing platform compared to the high fat diet plus stress induced group.

Though there was a marked increase in the body weight of the rats which were given high fat diet (90 days) but there was no change in the swimming ability of the rats (driving motivation) in the rats compared to the control group rats.

Statins, which are 3-hydroxy-methylglutaryl coenzyme A reductase (HMG-CoA reductase) inhibitors, have been clinically used for treatment of hyperlipidemia. It has been suggested that the pleiotropic effects of statins are responsible for their
neuroprotective action. Gavin J. Blake et al. stated that statins have anti-inflammatory properties[17]. Broncel M showed that statins have antioxidant properties and was useful in patients with dyslipidemia[18].

Parle et al. have reported that atorvastatin and simvastatin have improved the spatial learning in rats treated with scopolamine[19]. Wolzian et al. stated that, the prevalence of Alzheimer’s disease has been lowered in the patients taking lovastatin or pravastatin[20]. Rech et al. found that rosuvastatin reduced the oxidative damage in the hippocampus and improved short-term memory in rats that received iron in the neonatal period[21].

There was no difference in the neuroprotective properties between rosuvastatin (hydrophilic cannot cross the blood brain barrier) and atorvastatin (lipophilic)[22]. Recent studies on transgenic rats, zucker obese rats have shown that treatment with rosuvastatin have improved cerebral vascular responsiveness and they were restored to normal[23]. Present study on male wistar rats is in line with the previous studies that support the hypothesis that statins have neuroprotective effect[24-26].

Conclusion

Cure of cognitive disorders specially, dementias; is still a night mare in the field of medicine. Cholinesterase inhibitors, if used early in the course of disease can delay the progress of disease. Today, there is growing evidence that statins have several potential beneficial effects by mechanisms unrelated to changes of cholesterol metabolism. However, we still know very little about the clinical relevance of this lipid independent statin properties. This study, is establishing the advantage of controlling hyperlipidemia, which in turn protects from memory impairment and further broaden the use of lipid lowering agents.

References