Liver Regeneration: Summary of Involved Cell Types

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Abstract
The liver has unique capacity to regulate its growth and mass. Healing process in the liver is characterized by the proliferation of all existing cell lines within the liver, including hepatocytes, hepatic progenitor cells, non parenchymal epithelial progenitors and small hepatocyte like cells. Normally the liver exhibits a very low level of cell turnover, but when an abnormal hepatocyte loss occurs, a rapid regenerative response is elicited from all of these cell types in the liver. In this article, we review broadly the natural repair of the liver in response to injury.

Keywords: liver regeneration; Progenitor cells; Hepatocytes; Non parenchymal; Epithelial progenitors

Introduction
End-stage liver diseases, in particular cirrhotic liver, represent a worldwide health problem. Currently, liver transplantation is the only effective treatment, but it is associated with many problems, as, shortage of donors, operative damage, risk of immune rejection and high costs. Furthermore, liver transplantation may cause considerable long-term side effects, such as chronic renal failure, post-transplantation lympho-proliferative disorders and cardiovascular problems[1-2].

The emerging field of stem cell therapy has raised hope for improving treatment of liver diseases, because it is capable of promoting liver repair and regeneration with fewer complications. Stem cells are capable of self-renewal and can differentiate into specialized cell types. Cheng, (2008) has recently defined five minimal characteristics of stem cells, which includes self-renewal, maturation, resting mode, apoptosis and trafficking. The lack of any of these essential characteristics would make stem cells much less physiological in nature and not suitable for use as therapeutics[3].

Normally the liver has a very low level of cell turnover, but when an abnormal loss of hepatocytes occurs, a rapid regenerative response is elicited from all other cell types in the liver to restore the organ to its original state[4]. More severe forms of liver injury, particularly longstanding iterative injury (example: chronic viral hepatitis) or when replicative senescence is reached (example: steatohepatitis), activates a facultative stem cell compartment that resides within the intrahepatic biliary tree, giving rise to cords of bipotential transit amplifying cells (oval cells in rodents and hepatic progenitor cells in man) that can ultimately differentiate into hepatocytes and biliary epithelial cells[5].

A third population of stem cells with hepatic potential is located in the bone marrow; these stem cells usually make minimal contribution to regeneration but, after fusion with metabolically defective hepatocytes, they can be reprogrammed to contribute in a major way to restoring liver functions[6]. The natural repair of liver is primarily dependent on endogenous stem/progenitor cell pools (Figure 1), including hepatocytes, HPCs (hepatic progenitor cells) and OCs (oval cells)[7].
Hepatocytes

Hepatocytes are highly differentiated cells with different synthetic and metabolic functions. In addition to this, hepatocytes are the cells that normally carry the burden of regenerative growth after liver damage, so they are considered as the functional stem cells under most circumstances. In health, with individual life expectancy of over a year, hepatocytes are the only source of minimal ‘wear-and-tear’ renewal.

The regenerative capacity of the liver is particularly demonstrated when two thirds of the rat liver is resected by partial hepatectomy (PH), because the remaining part can re-grow to the original liver mass in about 10 days. This model has been deeply studied and has provided many data on the mechanisms involved in liver regeneration[4].

In response to such stimulus, all the normally quiescent hepatocytes leave G0 phase to enter the cell cycle under the influence of various growth factors. Hepatocyte proliferation is initiated in the perportal region of the liver and then spreads to the centrilobular region. This regenerative response requires each hepatocyte to undergo 1.4 rounds of replication on average, to restore normal liver size. Thus this means that hepatocytes have non limited replication potential. Hepatocyte transplantation trials in mice have shown that hepatocytes are capable of large-scale clonal expansion within a diseased liver.

Age has major influence on the response, as only in young rats (2 – 3 months old) do all hepatocytes enter the cell cycle at least once after a two-thirds PH, but in older rats (>2 years old) a significant number of hepatocytes do not respond and thus appear to have reproduction senescence[8]. After a two-thirds PH there is a latent period of about 15 h before the first perportal hepatocytes enter DNA synthesis, and the peak of the response is reached at 24 h, the hepatocyte DNA labeling index reaches 40%. After 4 days this hyperplastic response is effectively suspended and further increases in liver mass are achieved through hepatocyte hypertrophy. To retain correct cellular relationships, the non-parenchymal cells proliferate as well, although their kinetics of cell cycle entry lags a few hours behind that of hepatocytes[9].

Numerous growth factors and cytokines have been implicated in initiation and control of this regenerative response with the activation of various transcription factors, such as AP-1, NFkB and STAT-3[10]. To determine the actual regenerative potential of re-populating hepatocytes, Overturf and his colleagues performed serial transplantation of a limited number of unfractionated adult parenchymal hepatocytes into FAH (fumarylacetate-hydrolase)-deficient mice (a model of the metabolic liver disease tyrosinaemia type I). The results shows that hepatocytes could divide without loss of function at least 69 times[11].

Laconi and his colleagues blocked the proliferative potential of liver resident hepatocytes by using the pyrrolizidine alkaloid, retrorsine, after PH and then transplanted young adult DPPIV (dipeptidyl peptidase IV) F344 cells into DPPIV– F344 rats. They detected that in female rats 40 to 60% of hepatic mass was replaced within 1 year. Near-total replacement of hepatocyte mass was completed in male rats (98 to 99%) and lasted for 9 months after transplantation[12].

Oval cells (hepatic progenitor cells)

The liver has stem cells or HPCs that can be activated by persistent chronic liver[13,14]. The HPCs are the bipotent cells that can fully differentiate into hepatocytes and biliary epithelial cells[15]. Recent studies have shown that HPCs are present in the smallest and most peripheral branches of the biliary tree, the ductules and canals of Hering[16]. In rodents, adult HPCs are known as oval cells (OCs). The term ‘oval cells’ was first expressed by Farber, (1956) who found these cells with a characteristic morphological appearance after treatment of rats with carcinogenic agents, ethionine, 2-acetylaminoﬂuorine and 3-methyl-4-dimethyl aminobenzen[17].

These cells were described as ‘small oval shaped cells with scant lightly basophilic cytoplasm and pale blue-staining nuclei’[17]. Alison and his colleagues have shown that OCs is shown as a rich population of small round cells spreading from the periportal area and invading the parenchyma[5]. Other studies have also demonstrated that OCs are capable of generating either hepatocytes or biliary epithelial cells when hepatocytes are unable to mount a proliferative response to injury[18,19].

Non-parenchymal epithelial progenitor cells

The adult liver contains the multipotent NPE (non-parenchymal epithelial progenitor) cells[20]. In a recently published study by Duret and his colleagues, NPE cells were isolated from human liver tissue samples, located at a distance from the lesion, and cultured and characterized[20]. These cells expressed a markedly proliferative potential. They did not express the classic set
of stem/progenitor cell markers [Oct-4, Rex-1, AFP (α-fetoprotein), CD90, c-kit, and CD34] and were expressing albumin at low level.

When cultured at confluence in the presence of growth factors, example: HGF (hepatocyte growth factor) and either EGF (epidermal growth factor) or FGF4 (fibroblast growth factor 4), they entered a differentiation process toward hepatocytes.[20]

Their phenotypes were compared with that of mature human hepatocytes in primary culture. Differentiated NPE cells expressed albumin, α1- antitrypsin, fibrinogen, hepatobi- liary markers, such as cytokeratins 7, 8 and 19, liver-enriched transcription factors and genes characterized by either a fetal or a mature expression pattern. In addition, NPE cells could be isolated from the liver of different patients, irrespective of the absence or presence of lesions and differentiated toward hepatocyte-like cells with an intermediate hepatobiiliary and mature/ immature phenotype.[20].

These cells are likely to represent a resident progenitor cell population of the adult human liver. Dan and his associates have isolated a human fetal liver multipotent progenitor cell population that does not express liver-specific genes, but it is able to differentiate into functional hepatocytes and biliary epithelial cells. Moreover, these cells retain the multipotent potential to differentiate into fat, cartilage, bone and endothelial cells, and have a repopulation capacity in a liver injury mouse model[21].

Small hepatocyte-like progenitor cells

Another regenerative cell for liver injury is the SHPC (small hepatocyte-like progenitor cell). SHPCs share common phenotypes with hepatocytes, fetal hepatoblasts and OCs, but are phenotypically different. They express markers, such as albumin and immature form of AFP, and possess bile canalicu- li and store glycogen. If rats are exposed to retrorsine before PH, regeneration cannot occur from differentiated hepatocytes and the liver mass is restored by the induction and expansion of SHPCs.[22]. The origin of SHPCs is under debate. They may arise from a hepatocyte subpopulation that lack the required CYP (cytochrome P450) enzyme for retrorsine metabolism and therefore escape its mito-inhibitory effects.[22].

The results by Best and Coleman, (2007) provide direct proof that SHPC-mediated liver regeneration does not need OC activation or proliferation, and the SHPCs are not the progeny of OCs, but represent a distinct other population of liver progenitor cells.[23].

Conclusion

The combined evidence of these studies reveals that the liver has enormous replicative capacity in response to injury, it has adapted itself to the inflow of ingested toxins by the evolutionary development of unique regenerative properties and responds to tissue loss by the rapid division of mature cells. Pro- liferation of the parenchymal cells, i.e. hepatocytes and epithelial cells of the bile duct, and non parenchymal cells is involved in this cellular response. Resident hepatic stem/progenitor cells have also been identified in small numbers in normal liver and involved in liver tissue repair. Their potent role in the physiology, patho-physiology and therapy of the liver, however, is not yet precisely identified. Experimental analysis of the therapeutic value of these cells has been initiated but not fully understood. Cell based regenerative strategies aiming to replace, repair or enhance the biological function of damaged tissues or organs by utilizing their cells and bioactive molecules presents hope for patients especially in presence of transplantation problems and donors shortage.

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References

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