Neuronal Hypertrophy in Rat Colon Caused by Protein Deficiency

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Abstract
The effects of different protein levels associated with vitamin deprivation have been evaluated on tissues of high and low cell renewal. This study aimed to investigate the effects of available diet with 4% protein content associated with the maintenance of vitamin levels on the morphology of NADPH dp+ myenteric neurons of rats descending colon. Sixteen animals were randomly distributed into two groups, one that was fed with chow with 26% protein content (CG), and the other was fed with a 4% protein content diet (EG) during 12 weeks. Total preparations were used to make the myenteric plexus neurons of the descending colon evident when submitted to NADPH-diaphorase. The cellular bodies and nucleus of 300 neurons from each animal was measured. The diet with 4% protein and maintenance of vitamin and mineral content, similar to the ones found in normal protein diets, causes metabolic alterations that result in body weight loss. The myenteric neurons of the descending colon have non-significant reduction in the nucleus size whereas the cytoplasm presents a significant increase of the area. The importance of vitamin maintenance and the increase of the neuronal area are discussed as a possible compensating mechanism to keep neurotransmitter synthesis.

Keywords: Enteric nervous system; Protein malnutrition; NADPH-diaphorase; Large intestine; Myenteric plexus.

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Introduction
The enteric nervous system (ENS) consists of two main plexuses, the myenteric (Auerbach’s) plexus that is located between the longitudinal and circular layers of the muscular layer, and the submucosa (Meissner’s) plexus, located in the tela submucosa1,2. The main function of the myenteric plexus is the reflexive control of the intestinal motility. To perform its motor functions, the myenteric neurons can be subdivided into two large subpopulations: cholinergic and nitrergic with contractile and relaxing functions of the smooth muscle, respectively3. The nitrergic neurons have neuromediator nitric acid (NO)4 that is a neuroprotector, antioxidant and growth stimulant5,6. The myenteric plexus has morphological and functional characteristics that vary according to the different segments of the digestive tube in humans as well as in animals1,7, and it can have alterations in several experimental models like diabetes8,9, Chagas disease10,11, toxoplasma infection12,13 and protein malnutrition14-16. Protein deficiency causes decrease in the total neural population17-19 or increase in its density20,21. Myenteric neuronal subpopulations like cholinergic neurons22, positive NADPH-diaphorase19,16 and nitrergic neurons14,15 are also affected by malnutrition. Positive NADPH-diaphorase neurons are specially important to characterize nitrergic neuron population with inhibiting motor activity23,24. Previous studies have shown that these neurons can be affected by protein malnutrition and, therefore, this study aimed to evaluate the effects of severe protein malnutrition on the morphometry of myenteric nitrergic neurons of rats descending colon.

Material and Methods
All the experimental procedures were previously approved by the Ethics Committee for Research on Animals Experimentation (CEPEEA) of Universidade Paranaense. Sixteen male, 90-day-old, Wistar rats (Rattus norvegicus) (body mass of 313.92 ± 9.79g) were separated into a control group (CG: n=8), fed normally, and an experimental group (EG: n=8), submitted to protein malnutrition. They were kept in individual cages in an animal house with controlled temperature and humidity and a 12-hour light/dark cycle.

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CG was fed NUVILAB® added with 26% of protein, and the EG was fed a diet prepared with 4% of protein through the addition of corn starch and keep the vitamin and mineral balance according to the description by Araujo et al.[25]. Both groups were given chow and water ad libitum for 90 days. After this period, the rats from both groups were anesthetized and euthanized with Acepran (1.26 mL/Kg), Ketamine-10% (1.26 mL/Kg), Xylazine-2% (0.42 mL/Kg) and Atropine-1% (0.22 mL/Kg) intramuscularly[26].

Descending colon was histochemically submitted to NADPH-d to make it evident. A segment of each animal was removed, washed in phosphate buffer solution (PBS pH 7.4), immersed and fixated in paraphormaldehyde at 4%1, prepared in pH 7.4 0.1M for 30 minutes; next, they were immersed in a solution of Triton X-100® 0.3% dissolved in PBS pH 7.4 0.1M for 10 minutes and posteriorly washed 10 times (10 minutes each) in PBS pH 7.4 0.01M. Then, to make neuronal β-NADPH+ evident, they were immersed in an incubation medium containing a solution of 0.05 de Nitroblue Tetrazolium NBT2 in 200 mL of tris-HCl buffer solution; 0.1 g of β-NADPH3 and 0.6 mL of Triton X-100. The segments were washed in PBS 0.01M pH 7.4 (three times, 5 minutes each), and the reaction interruption occurred by adding paraformaldehyde at 4% in PBS 0.01M pH 7.4 [24].

For the morphometric analysis, the area of the cell and nucleus body of 100 neurons of each region of the intestinal circumference (mesoelastic, intermediate and antimesoelastic) of all animals from both groups were measured through the images obtained by a 40x lens microscope with a video camera coupled to a computer with an image analysis software4. The cytoplasm area was calculated from the differences in these areas. The ratio between the nucleus area and the cell body area, as well as their correlation, was calculated. The neurons were classified in 50 µm² intervals for the cell body.

Initially, data was submitted to D’Agostino-Pearson’s test or Shapiro-Wilk’s test to verify distribution. Data for normal distribution are presented as means ± standard deviation. In this case, to compare the groups, Student’s t test was utilized for the independent samples. Data with free distribution are presented as means and percentages (25:75), therefore, the comparison between the groups was done by Mann-Whitney’s test. The correlation analysis was verified with Sperman’s non-parametric test. In all statistical tests, p values lower than 0.05 were considered significant. All the analyses were performed using BioEstat 5.0 software[27].

### Results

The body mass in the beginning of the experiment was 311.83 ± 11.27g in CG and 316 ± 9.99g in EG (p > 0.05). In the end, the body weight was 403.43 ± 11.25 g in CG and 246.50 ± 18.34g in EG, (p < 0.05).

There was an increase in the cell body area, cytoplasm area, cell body/nucleus area ratio without alteration of the nucleus in EG (Table1).

<table>
<thead>
<tr>
<th>Measurement</th>
<th>CG</th>
<th>EG</th>
</tr>
</thead>
<tbody>
<tr>
<td>Cell body (µm²)</td>
<td>181.30 (144.00: 218.72)</td>
<td>193.71 (159.22: 232.14)*</td>
</tr>
<tr>
<td>Cytoplasm (µm²)</td>
<td>140.13 (108.67: 173.36)</td>
<td>156.25 (124.50: 192.07)*</td>
</tr>
<tr>
<td>Nucleus (µm²)</td>
<td>39.00 (28.66: 48.52)</td>
<td>36.86 (28.66: 45.99)</td>
</tr>
<tr>
<td>Ratio</td>
<td>0.21 (0.17: 0.26)</td>
<td>0.19 (0.15: 0.23)*</td>
</tr>
</tbody>
</table>

Values marked with an asterisk (*) in the same line presented significant difference (p < 0.0001).

The correlation degree between the body cell area, nucleus area and cytoplasm area of positive NADPH-diaphorase myenteric neurons (NADPH-d+) of the descending colon is shown in Figure 1 for CG and in Figure 2 for EG.

Figure 1: Dispersion diagram of the correlation between the cell body area and the nucleus area (A, r = 0.59, p<0.05), body cell area and cytoplasm area (B, r = 0.97, p<0.05) and cytoplasm area and nucleus area (C, r=0.38, p<0.05) of NADPH-diaphorase myenteric neurons of descending colon of normally fed rats (Control Group).

Figure 2: Dispersion diagram of the correlation between the cell body area and the nucleus area (A, r = 0.53, p<0.05), body cell area and cytoplasm area (B, r = 0.97, p<0.05) and cytoplasm area and nucleus area (C, r = 0.32, p<0.05) of NADPH-diaphorase myenteric neurons of descending colon of rats submitted to protein malnutrition (Experimental Group).

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1 Merck, Damstadt, Germany
2 Nitroblue Tetrazolium Giboibrl, New York, USA
3 Sigma, Steinheim, Germany
4 Motic Imagem Plus versão 2.0
The number of distributed neurons in classes, according to the body cell area, as well as the body cell area and nucleus area ratio, is show in Figure 3.

Figure 3: Distribution of myenteric neurons of descending column of control rats (control group) stained by NADPH-diaphorase and submitted to protein malnutrition (experimental group) distributed according to the classes of the cell body area. The values do not present significant difference p>0.05.

The protein malnutrition increased the cell body area due to the augmentation of the cytoplasm volume without the corresponding increase in the nucleus volume.

Discussion

The protein content of 4% in the chow through the addition of cornstarch was insufficient to allow the animals’ body mass development. The body mass of EG animals was 38.89% lower than the CG, besides not gaining weight EG rats had a 22.24% body mass decrease after 12 weeks of treatment. Reduction of the body mass was not found in rats that were fed with 8% protein content diets[13], on the other hand, rats that received diets with less than 5% protein content, similarly to the results found in this study, lost weight[14]. This shows that when the protein ingestion is very low, the organism has difficulty to transform carbohydrates into fat reserve, possibly due to the lack of aminoacids for the synthesis of structural and functional proteins like enzymes that are related to the conversion of glycides into lipids. On the other hand, the reduction in weight indicates the use of fat stores in the processes of energy production and possible insufficiency of protein for maintenance of muscle mass. Araujo et al.[14] points out that a hypoprotein diet for a long period triggers a mechanism that allows adaptation when there is low protein availability in order to preserve the nervous tissue.

In this study, an increase of the body cell area of NADPH-d neurons in EG was observed and it may be due to the increase of the cytoplasm volume, once the nucleus had non-significant reduction of its size.

The increase of the cytoplasm area can be relate to different mechanisms: (1) due to neuronal loss related to diet with 4% proteins[14], the remaining neurons, possibly increase the volume of the cell body as part of an adaptive mechanism to compensate the lost neurons through an increase in their cytoplasm system for protein synthesis, assuring the maintenance of the target tissue. Zanoni et al.[28] report that the neuronal hypertrophy in diabetic rats is associated with the attempt to keep the neurotransmitter production in favor of the lost neuronal population; (2) would the augmentation of the neuronal body be the initial stage of apoptosis? The protein restriction can induce metabolic failure and result in organnelle damages, mainly in mitochondria. Mitochondrial lesion activates the intrinsic pathway to apoptosis by means of caspases. Thus, it is possible to infer that the lower protein content in the diet caused the deprivation of neuronal growth factors (NGF) and other essential substances that regulate the bioenergetics flow, resulting in the activation of pro-apoptotic cytoplasm proteins of Bel-2 family[29]. The increase of the cytoplasm volume associated with the 5.6% reduction (p<0.0001) of the nucleus area corroborate this hypothesis because this morphological aspect is common in cells in the beginning of apoptosis. With the evolution of apoptosis, the nucleus becomes disproportionately small (pycnotic nucleus) and with intensely condensed chromatin. This can be visualized by staining techniques that use acid/basic differentiation[30]; (3) would hypertrophy observed in this study be due to an attempt to keep the NADPH-d enzyme expression? Similarly to this study, a previous study with the same animals was associated to the myenteric neuronal death[14], indicating a probable increase in the activity of the nitric oxide synthesis (NOS) and in the production of nitric oxide (NO) in the remaining neurons.

Our results suggest that the vitamin and mineral maintenance utilized in this study prevented the atrophy of NA-DPH-d neurons; in addition, it augmented the cytoplasm volume which can be a consequence of the increase of the synthesis of the remaining neurons to compensate possible neuronal losses and keep the neurotransmitter levels compatible to the necessities of the target tissue. The intensification of neuronal damage by the association of the vitamin deficiency with the protein deficiency was evident in a previous study[15] carried out with rats that were fed with 4% protein content diets and reduced vitamin levels, resulting in neuronal loss and atrophy of remaining nitrergic neurons of the myenteric plexus of the descending colon. In the cecum, Muniz et al.[31] verified that there was a reduction in the area of the body cell and nucleus of NADPH-d neurons of animals submitted to a hypoproteic diet (4%) for 90 days; however, with the preservation of the neuronal population, this atrophy occurs by neuronal modulation when there is a lower amino acid availability for protein synthesis. These findings are divergent from the ones found in this study, and suggest that there may be different adaptive mechanisms to protein deficiency, among them the maintenance of neuron amount associated with the reduction of the cellular body volume, and consequent reduction of the neuron synthesis machinery. Another possibility is the reduction of the amount of NADPH – dp’ neurons, as verified in the descending colon of rats in a previous study[14] with the same diet utilized in this study. In that case, the increase of the cellular body volume, found in the present study, could be a consequence of the increment of the remaining neuron synthesis. The neurons made evident by NADPH-d histochrometry represent a neuronal subpopulation involved with the nitrergic pathway that has inhibiting potential to the intestinal smooth muscle contraction[24,25]. Therefore, hypertrophy, verified in this study, allows us to infer that there was an alternation in NADPH-d enzyme expression, resulting in a neurotransmitter unbalance of descending colon and that this can be related to the diarrhea observed along the experimental period[14].

In this experiment, it was observed that the positive correlation between the body cell and the nucleus of nitrergic neurons was 59% in CG and 53% in EG, suggesting that the growth and/or reduction of the nucleus size did not follow the cell body size. That was not found when the correlation between the body and cytoplasm was observed (97%), indicating that the cell body of myenteric neurons can follow (as a cause or a consequence) the alterations in the cytoplasm of the same neurons.
When analyzing the distribution of the cell body area, it was observed that there was no difference between the analyzed classes. Despite the homogenous distribution, the neurons of both groups were mainly from 151 to 200µm². Regarding the cell body area, the neuron incidence between the different classes did not present a significant difference between the groups; there was a predominance of neurons whose nucleus occupies from 16 to 20% of the cell body in both groups, indicating that malnutrition kept distribution uniformity of the studied classes, even though it caused hypertrophy.

To sum up, the provision of the diet with reduced protein content to 4% through the addition of starch associated with the maintenance of the vitamin and mineral levels, similar to the one found in normal protein diets, causes the metabolic alterations that result in body weight loss that is possibly related to the lack of amino acids for the synthesis of enzymes related to the utilization of carbohydrates as an energy source and the conversion of fat reserves. In NADPH–dp+ neurons of the descending colon there is a non-significant reduction of the nucleus size while the cytoplasm increases its area significantly.

References