Water \( H_2O_2 \) Levels as Factor in Swimmers Melanoma

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Abstract:

Introduction: This study proposes a foundation which could demonstrate a published \( H_2O_2 \) hypothesis of cancerogenesis. A mechanism explaining the high incidence of melanoma in swimmers is described.

Methods: Human hairs immersed in drops of pure water with resistivity of 18.2 MΩ × cm (million ohms) were in indirect contact with adjacent drops of 35% \( H_2O_2 \). Digital microphotographs and video-recordings were obtained for further analysis.

Results: This approach allows visualization of details that could not be analyzed previously. The findings show that \( H_2O_2 \) penetrates hair follicles either at the sites of injury to external structures or at the intact shaft/skin junction.

Conclusions: In hairs immersed in pure water mixed with a low concentration of \( H_2O_2 \), sebum and contiguous dermal sheaths blocked exogenous \( H_2O_2 \). Conversely, in injured hair follicles and at the intact shaft/skin junction, \( H_2O_2 \) penetrated the tissue and was subsequently decomposed by catalase. This mechanism is proposed for the high incidence of Swimmers Melanoma.

Keywords: \( H_2O_2 \), Oxidative Stress, Swimmers Melanoma, Pig Melanoma Formation, Biophysics cancerogenesis

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Abbreviations: Decomposition: Breaking down of a molecule; \( H_2O_2 \): Hydrogen Peroxide; RH: Relative Humidity- Measure of ambient water vapor; ROS: Reactive Oxygen Species; 18.2 MΩ × cm (million ohms): Pure water resistivity. This only accounts for the dissolved ionic impurities commonly found in water; Substrate: The substance acted upon by an enzyme. In this manuscript \( H_2O_2 \). Enzyme activity is directly proportional to substrate concentration; DOM: Dissolved organic matter in fresh water. UV Excitation: UV portion of sunlight (limited to depth of UV in body of water).
Materials and Methods

Glass slides (25×75×1 mm, #1301) were purchased from Globe Scientific Inc. Certified food grade H₂O₂ (35%) and bovine liver liquid catalase (purified, thymol-free) was purchased from Sigma-Aldrich (StockC-40, one gram 11000 units). Very pure water with resistivity of 18.2 MΩ × cm (million 112 ohms) supplied by The University of Oklahoma, Health Sciences Center was used. The equipment included a model OS425-LS non-contact infrared thermometer, Celestron model 44348 digital video microscope, 117 Acurite model 01538CDI remote weather station, Apple MacBook computer, and Apple Inc. iPhoto 8.1.2 application.

Harvesting and mounting of hair samples

Sixteen hairs were plucked from the scalp of the author using tweezers. Care was taken to select hairs with large visible roots. Using a fresh double-edge razor blade, ten of the hair follicles were gently injured after placing on a dry slide. This maneuver produced four partially transected follicles with different follicular injury patterns. The individual hairs (12 intact and 4 injured) were individually placed on the center of clean glass slides then covered by two drops of very pure water with a resistivity of 18.2 MΩ × cm. Two to three drops of 35% H₂O₂ were delivered via a micropipette to the same slide 4 to 5 mm distant from the pure water drops. Care was taken not to mix the drops. Each preparation was mounted on a digital video-microscope platform and observed for any changes of the hair follicles. Gas bubbles emanating from the hair follicles appeared slowly on the hair tissue. Once the slow bubbling started after approximately 8 ± 3 minutes (Fig 1A), microphotographs and video-recordings were obtained and the data were stored for subsequent analysis. Ancillary testing demonstrated the gradual transfer of H₂O₂ into the pure water drops.

Results

All intact follicles showed bubbling activity; it occurred near the shaft/skin boundary in 12 follicles (Figure 2 & 3) and Supplementary Video #0043) and at the bulb and suprabulbar external areas in the four injured follicles. In all the injured follicles, O₂ gas was observed to be flowing slowly between the cortex and medulla, exiting at the injury site (Figure 4 & 5).
Swimmers Melanoma and H$_2$O$_2$

Discussion

Oxidative stress due to tissue metabolism in hair follicles has been previously demonstrated$^{[11,12]}$. Hair follicles produce catalase in order to decompose ROS and achieve homeostasis. The present findings were possibly due to the very slow bubbling observed during the endogenous catalase-mediated decomposition of H$_2$O$_2$ that penetrated the hair follicles.

The first thought was that the presence of catalase in the scalp$^{[13]}$ could be the cause of the intense bubbling observed at the shaft/skin junction site. High magnification views showed that the bubbling originated away from the shaft/skin boundary surface. Since the shaft/skin junction area was uninjured, the observations prompted the consideration of whether the shaft/skin junction area in the skin was a point of spontaneous entry for H$_2$O$_2$.

Sebum/Dermal sheaths as barriers

Observed was that exogenous H$_2$O$_2$ penetrated deep into hair follicles. In seeking a mechanism for this observation, it was noted that when hair follicles were injured, the protective sebum coat and dermal sheaths were compromised (Figure 6). This would allow the exogenous H$_2$O$_2$ to penetrate the internal tissue layers where it would be decomposed by the ubiquitous enzyme catalase.

The present study also presents an association of sebum combined with intact dermal sheaths acting as a barrier to ROS external toxicity.

Summary and Conclusions

Swimmers melanoma

Two decades ago, it was postulated that the recreational exposure to sunlight did not fully explain the current trends in melanoma incidence$^{[14]}$. The authors suggested that the “positive association between a history of swimming and melanoma risk suggests that carcinogenic agents in water, possibly chlorination by products, play a role in melanoma aetiology”. Additionally, considering the skin dryness of swimmers, the same authors stated that this is “caused by a combination of the dilution of natural sebum and by the osmotic gradient produced when the body is immersed in water, drawing hydration from the outer skin layers.” Another study documented several dermatological problems related to prolonged or repetitive water immersion, including skin denuding of the protective sebum coat$^{[15]}$. The protective barrier function of the skin against ROS has been identified previously$^{[16]}$.

The experimental documentation presented in this manuscript supports the view that H$_2$O$_2$ fuels aging, inflammation, cancer metabolism, and metastasis$^{[17]}$. Furthermore, the present observations provide a basis for testing the hypothesis that ROS reactions are associated with cancerogenesis$^{[18]}$. We demonstrated that in hair follicles, compromised sebum/dermal sheath layers and uncompromised shaft/skin junction allow H$_2$O$_2$ penetra-
tion, resulting in repetitive ROS reactions and possible initiation of diseases. This could include melanoma.

Addendum

The data presented in this manuscript also supports that in fresh or seawater, the penetration of ROS into the submerged hairs occurs at the hair shaft/skin interface, as well as through the injured external structures. The risk for "swimmers melanoma" could be explained as follows: “The formation of hydrogen peroxide results principally from the UV portion of sunlight exciting humic substances in the water and thereby leads to the formation of superoxide ion, which reacts with itself to form H$_2$O$_2$. Because this production is limited to the depth of UV light penetration, its vertical distribution provides a sensitive tracer for mixing processes”[19].

Factor affecting the formation of H$_2$O$_2$ in fresh waters

DOM $\rightarrow$ UV Excitation $\rightarrow$ O$_2$ $\rightarrow$ H$_2$O$_2$

* DOM = Dissolved Organic Matter
** UV Excitation = UV portion of sunlight (limited to depth of UV in body of water)
*** O$_2$ $\rightarrow$ UV portion leads to formation of superoxide ion, which reacts with itself to form H$_2$O$_2$. [20,21]
**** H$_2$O$_2$ = Vertical distribution of Hydrogen Peroxide molecules now in body of water.

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References


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