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Comparative between Agno₃ and Kmno₄ as an Alternate Method of Castration in Lwy Boar

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Abstract

The study aims to determine the efficiency of castration between two different chemicals for castration. The study was carried out in 16 Large White Yorkshire male pigs of 2-3 months and reared till slaughter age. Intratesticular injection of chemical was done when the pig reached the age of 2-3 months for inducing castration in the pig. The animals were divided into two groups consisting of 8 animals in each group, in which in Treatment-1 or T_1 the pigs were castrated chemically using 5% AgNO₃ and for the other group Treatment-2 or T_2 were castrated using KMnO₄. Statistical analysis revealed that there was no significant (P>0.05) difference between the two groups in testicular measurement and the testosterone concentration. But, numerical observation for both of the findings reveals that there was a different between the two treatment groups with lower testicular measurement and testosterone concentration in T_2 group. Histopathologically the testes in T_2 group revealed lesser number of sertoli cells in the testicular tissue and most of the seminiferous tubules were devoid of spermatozoa. From the present finding it may be concluded that castration using KMnO₄ may be the best option for castration of the pigs.

Introduction

Livestock plays an important role for increasing the production of animal origin food like milk, meat and eggs as well as Socio Economic Development of the state. Pork is highly accepted meat in India and abroad. Pork of castrated pig is also preferred to un-castrated one due to the presence of boar taint in the meat. Castration was done mainly when the pigs reached the age of 2-3 months of age^[1]. Breeding boars were also castrated under general anaesthesia after they had become unproductive castrations were performed to decrease the boar taint which is an unpleasant odour which is pronounced upon heating of pork, mainly entire males. The traditional surgical method of castration in pig is highly technical, costly and difficult to adopt by common pig farmer^[2-4]. In addition, the physiological reactions and behaviours of castrated pigs are also modified^[5]. Thus, this experiment was planned to evolve a suitable technology of chemical castration in pigs for reducing the boar taint and also to prevent excessive stress to the animals.

Materials and Method

The study was carried out at Instructional Livestock Farm Complex College of Veterinary Sciences & Animal Husbandry, Central Agricultural University, C.A.U, Selesih, Aizawl, Mizoram, India. A total of 16 24 Large White Yorkshire (LWY) of male pigs of 2-3 months of age were divided into 2 groups of 8 animals each. In Treatment 1 (T_1), chemical method of castration was conducted by injecting 2 ml of 5% Silver Nitrate Solution intratesticularly as per Kang *et al.* (1993)^[6] and in Treatment 2 (T_2) chemical method of castration was conducted by injecting 2 ml of 0.25 g potassium permanganate + 17 ml glacial acetic acid + 83 ml distilled water intratesticularly as per Giri *et al.* (2002)^[7]. Administration of chemicals was done by sharp needle and syringe pushed through epididymis into the body of testis to discharge the fluid from one end to the other in a withdrawing fashion. Animals of both groups were maintained on uniform feed and managemental condition up to 180^{th} day of castration. Testicular length, breadth and depth of pigs were recorded at monthly intervals. Blood from the pigs were taken at monthly interval and serums were separated for testing the testosterone

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concentration. Histopathological study of the testis recovered from chemically castrated group was conducted to find out the changes in the testis. Finally statistical analysis was done using SPSS package 20.0

Results and Discussion

Testicular measurement (mm)

The mean \pm SE of testicular measurement (mm) of LWY male pigs at 2 month of age are presented in Table 1. There were no significant differences between the treatment group in testicular measurement, the mean measurement of the testis for the length, breadth and depth under the treatment T₁ and T₂ were 38.00 \pm $0.58 \& 38.93 \pm 0.37$; $15.97 \pm 0.49 \& 16.67 \pm 0.17$ and 14.50 \pm 0.36 & 14.6 \pm 0.87 mm respectively. In T₁ it was found that there was an increasing trend in the length, breadth and depth of the testicles till the end of the rearing period i.e. 180 days, but the final measurement in the present study was found to be lesser than the finding of Singh et al. (2018)[7] who had done the gross morphological study of the testes in LWY intact boars. The increase in the size of the testes from 4^{th} months of age in T, may be due to regeneration of the testicular tissue in the later stage of the rearing period. The increase in the size of the testes may be the reason for increase in the concentration of the testosterone hormone, as the size of testes is correlated with the amount of testosterone production, which was in support by the findings of Bekaert et al. (2012)^[8] and Bernau et al. (2018)^[9,10]. Whereas, in T₂ that there were decreased in the size of the testes from 4 months to 5 months of age and from 6 months the testicle was complete atrophied hence, measurement was not possible. This finding is similar to the finding of Giri et al. (2002)^[7] and Kang et al. (1993)^[6] who had observed that the testicles had completely atrophied in chemical castration and later act as a rudimentary organ in pig later stage of life.

 Table 1: Mean Testicular Measurement between the Different Treatments at age (mm)

Age (Mon-	M e a - surement	Treatment group			
ths)	(mm)	T1	T2	t value	p Value
2	Length	$38.00{\pm}~0.58$	38.93 ± 0.37	1.06 ^{NS}	0.10
	Breadth	$15.97{\pm}0.49$	16.67 ± 0.17	0.95 ^{NS}	0.52
	Depth	$14.50{\pm}~0.36$	14.6 ± 0.87	0.04 ^{NS}	0.66
3	Length	39.67 ± 0.33	40.83 ± 0.44	2.11 NS	0.10
	Breadth	$17.33 \ \pm 0.33$	$18.57\ \pm 0.35$	2.57 ^{NS}	0.62
	Depth	16.00 ± 0.58	16.27 ± 1.11	0.19 ^{NS}	0.56
4	Length	55.00 ± 5.00	31.47 ± 10.77	2.00 NS	0.12
	Breadth	56.63 ± 1.26	14.67 ± 6.12	1.68 ^{NS}	0.17
	Depth	19.13 ± 0.38	11.40 ± 3.80	2.02 ^{NS}	0.11
5	Length	72.67 ± 3.93	27.10 ± 16.95	2.43 ^{NS}	0.13
	Breadth	28.97 ± 1.93	$12.50\ \pm 9.65$	1.78 ^{NS}	0.21
	Depth	21.80 ± 0.46	8.97 ± 6.27	2.03 ^{NS}	0.11

(*) Significant (P \leq 0.05), (**) Significant (P \leq 0.01) and NS Non-significant

Note: Means bearing at least one common superscript in each row do not differ significantly.

Testosterone Concentration (pg/ml)

The mean testosterone concentrations (pg/ml) for T₁ and T₂ at 6 months age were 2148.17±604.06 and 1879.33±252.70 pg/ml as shown in Table 2. Statistical analysis shows a non-significant difference between the groups, which might be due to the high standard deviation which may result in non applicable to statistical treatment. The concentration of testosterone level in T, and T₂ group though reduced at 3 months was found to increase subsequently. The present finding was in contrast with the findings of Fahim (1994)^[11] (7) who had reported that testosterone concentration decreases at 22 weeks of age when animals was injected with 100 mg of zinc arginine intratesticularly in pigs. Same findings was also reported in other species of animals by Vanderstichel et al. (2014)^[12] in dogs by using different chemicals; Al-Asadi & Al-Kadi (2012)^[13] (1) in bucks after chemical castration; Canpolat et al. (2006)^[14] and Cohen et al. (1991)^[15-18] in bulls. The variations in the findings may be due to partial destruction of the testes, or especially due to the partial immunity of the interstitial cells being destroyed. It might also be due to the fact that there was a regeneration of the testicular tissue from the 5th months of age, which led to higher concentration of the hormones testosterone.

 Table 2: Mean testosterone concentration between the different treatments at different age. (Pg/ml)

Age	Treatment group				
(Months)	С	T1	T2	F value	P value
2	1574.13± 884.62	1006.07± 220.12	1145.10± 126.55	0.16 ^{NS}	0.85
3	$\begin{array}{c} 1293.5 \pm \\ 420.49 \end{array}$	$\begin{array}{r} 886.07 \pm \\ 272.93 \end{array}$	934.13 ± 63.48	0.49 ^{NS}	0.63
4	926.83 ± 135.53	952.73 ± 207.87	$\begin{array}{r} 867.47 \hspace{0.1 cm} \pm \\ \hspace{0.1 cm} 4.04 \end{array}$	0.56 ^{NS}	0.60
5	$\begin{array}{r} 860.17 \pm \\ 102.481 \end{array}$	1119.40± 117.01	997.50 ± 31.75	2.00 ^{NS}	0.22
6	$\begin{array}{r} 800.80 \pm \\ 436.69 \end{array}$	2148.17± 604.06	1879.33 ±252.70	2.79 ^{NS}	0.14

(*) Significant (P \leq 0.05), (**) Significant (P \leq 0.01) and NS Non-significant

Note: Means bearing at least one common superscript in each row do not differ significantly.

Histopathology

The present study reveals that in T_1 treatment group there was a remarkable increased population of interstitial cells, seminiferous tubules containing a tuft of developing spermatozoa with tails and sertoli cells in the tubules. But no mature spermatozoa could be seen in the centre of the seminiferous tubules.

While in T_2 the testicular tissue showed a remarkable increased in the population of interstitial cells but the tubules were vacuolated with no spermatozoa. The findings were in accordance with the report of Giri *et al.* (2002)^[7].

The histopathological study of the testes of T_2 group reveals that due to lesser number of sertoli cells in the testicular tissue and most of the seminiferous tubules were devoid of spermatozoa. The interstitial cells showed hypertrophy giving an adenoid appearance (Figure.1) with necrobiotic changes in Citation: Vanlahmangaihsanga, L., et al. Comparative between Agno3 and Kmno4 as an Alternate Method of Castration in Lwy Boar. (2020) J Vet Sci Animal Welf 4(1): 10-13.

few cells. The present findings in T_1 and T_2 may be concluded that due to irritation caused by chemical castration, the seminiferous tubules were devoid of spermatozoa and there was disappearance of germinal cells due to its delicate structure and high sensitivity to chemicals. However, the interstitial cells being less sensitive to such chemicals got better nourished in post inflammatory period and become hypertrophied with adenomatous appearance, which might be due to continued secretion of androgen (Giri *et al.*, 2002)^[7].



Figure 1: (a) Cross section of testicle in pigs under chemical castration with $AgNO_3$ solution (T₁).

(b) Cross section of testicle of pig under chemical castration with $KMNO_4$ solution (T₂). (Testis H & E 400X)

Conclusion

From the present study statistical analysis reveals no significant difference between the treatment groups, but numerical values reveals that in T_2 groups there were lower testicular measurement and testosterone concentration. Further, a T_2 group shows a significant change in the histopathological morphology. It may thus be concluded that chemical castration by KMnO₄ may be a better method in castration of the pigs. But, further research may be carried out on chemical castration for longer rearing period to ascertain the present finding.

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